



Xanthine oxidoreductase is central to the evolution and function of the innate immune system

Claudia Vorbach¹, Roger Harrison² and Mario R. Capecchi³

¹Ontario Cancer Institute, Princess Margaret Hospital/University Health Network, Toronto, Ontario, M5G 2C1, Canada

²Department of Biology and Biochemistry, University of Bath, Bath, BA2 7AY, UK

³Howard Hughes Medical Institute, Department of Human Genetics, University of Utah, Salt Lake City, UT 84112, USA

The housekeeping enzyme xanthine oxidoreductase (XOR) has been studied intensively over the past 100 years, yet the complexity of its *in vivo* function is still poorly understood. A large body of literature focuses on the different catalytic forms of XOR and their importance in the synthesis of reactive oxygen and reactive nitrogen species, which are involved in many disease processes. More recently, various protective physiological roles of XOR have been recognized. We summarize for the first time that XOR is a component of the innate immune system. Because XOR is involved in multiple features of innate immunity we suggest that it is central to the evolution and function of this ancient defense system. We present evidence suggesting that XOR is a direct downstream target of the evolutionarily conserved Toll-like receptor–NF- κ B pathway and discuss that numerous forms of post-translational modification of XOR could provide intrinsic molecular switches that make XOR an ideal component of various fast innate immune responses.

Xanthine oxidoreductase (XOR) is best known as an evolutionarily conserved housekeeping enzyme with a principal role in purine catabolism [1,2]. Humans with mutations in *XOR* suffer from xanthinuria, an autosomal-recessive disorder resulting in kidney stone formation and urinary tract disorders [3]. By generating mice with a targeted disruption of *XOR*, we discovered the additional role of XOR as an essential protein for milk fat droplet secretion from the lactating mammary gland [4]. This surprising function prompted us to review the complex role of XOR in mammals and throughout the animal kingdom. There is increasing evidence that XOR has additional physiological functions associated with its synthesis of reactive oxygen species (ROS) and reactive nitrogen species (RNS), which have important roles in inflammation and host defense [5,6]. Here, we propose for the first time that XOR is an evolutionarily conserved component of the innate immune system.

XOR performs various cellular protective functions central to the innate immune system

In purine catabolism, XOR catalyzes the oxidative hydroxylation of hypoxanthine to xanthine and subsequently of xanthine to uric acid [1,2]. This, and presumably other, metabolic reactions of XOR have a far-reaching impact on cellular homeostasis, cellular protection from toxic compounds and also systemic protection, known as innate immunity. Uric acid and its oxidation product allantoin act as potent antioxidants and free radical scavengers, necessary to protect a cell from oxidative damage caused by numerous naturally occurring ROS and RNS [7–9]. XOR has, therefore, important functions as a cellular defense enzyme against oxidative stress. Interestingly, XOR itself contributes to the synthesis of numerous ROS and RNS [1,2,5,6]. XOR exists in two forms, as xanthine dehydrogenase (XD; EC 1.1.1.204), which is the primary gene product of XOR, and as xanthine oxidase (XO; EC 1.1.3.22), which is formed through post-translational modifications of XD. XD favors the cofactor NAD⁺ as its primary electron acceptor, yet XO is unable to bind NAD⁺ and uses O₂ as its electron acceptor. With both forms, but particularly with the XO form, numerous ROS and RNS are synthesized [1,2,5,6]. Consequently, the synthesis of both an antioxidant (uric acid) and numerous free radicals (ROS and RNS) makes XOR an important protective regulator of the cellular redox potential [10]. Moreover, XOR is also a potent detoxification enzyme, probably owing to its multifunctional enzymatic activities [11]. This detoxification function is not restricted to XOR but is rather generally found in the molybdopterin family of enzymes, including aldehyde oxidase (AO) [11]. In this context, AO is of particular interest because its amino acid identity and its intron–exon structure is similar to XOR, suggesting that both enzymes evolved through gene duplications [12]. Not surprisingly, XOR and AO have a large number of common substrates and similar expression patterns [12].

The innate immune system is an evolutionarily conserved, rapid, non-specific first-line defense mechanism that incorporates elements of cellular differentiation and protection against oxidative stress [13]. Innate immunity is composed of: (i) surface epithelia that provide local physical and molecular barriers, (ii) inflammatory

Corresponding author: Mario R. Capecchi (mario.capecchi@genetics.utah.edu).

reactions and the activation of conserved cell-signaling pathways, (iii) numerous systemic protective molecules and (iv) various phagocytotic cells. All of these components work together to resist and prevent the action of toxic molecules and the rapid spread of potentially fatal pathogens. The protective functions of XOR in innate immunity are, as at the cellular level, linked to its detoxification reactions, its synthesis of uric acid and, particularly, its synthesis of numerous ROS and RNS.

Uric acid not only acts as a cellular but also as a systemic antioxidant and free radical scavenger, besides being an anti-inflammatory effector with numerous protective roles in the body [14–16]. ROS and RNS perform, at low levels, numerous cellular and physiological functions as second messengers but, at high levels, can act as microbicidals [17–19] (Fig. 1). XOR generates the ROS superoxide anions (O_2^-), hydroxyl radicals (OH^\cdot) and hydrogen peroxide (H_2O_2) [20]. Those ROS can generate an entire cascade of microbicidal reactions and products by participating in the synthesis of additional antimicrobial and cytotoxic molecules, thereby broadening their range of action [21]. RNS derive from the free radical nitric oxide (NO), a product of NO synthase as well as XOR [22]. NO itself is antimicrobial and cytotoxic, and it is further involved in the generation of an array of reactive molecules and even more potent antimicrobial substances, which makes NO a defensive molecule against various pathogens, tumor cells and alloantigens [23,24]. XO has also been implicated in protective antiviral responses by catalyzing the conversion of retinaldehyde to retinoic acid. Derivatives of retinoic acid can inhibit viral replication, thus potentially preventing the spread of viral diseases [25]. As we summarize here, XOR and its products are directly and/or indirectly involved in multiple features of innate immunity in mammals and, furthermore,

have a conserved defense function throughout the animal kingdom.

XOR is expressed and secreted by numerous surface epithelia

Because most pathogens enter the body through surface membranes, epithelial tissues are an important component of the non-specific immune system. Surface epithelia have evolved many defense mechanisms, among which are the expression and secretion of numerous molecules. These molecules perform broad-spectrum antimicrobial functions by neutralizing and inactivating pathogens and/or by generating transient microbicidal and cytotoxic substances, such as ROS and RNS. As a housekeeping enzyme, XOR is presumably expressed in all cells, yet, high levels of constitutively expressed XOR, as well as AO, are found in many mammalian epithelial and capillary endothelial tissues of various organs [26–29]. The intensity of XOR activity appears to be organ-dependent [28,29]. Numerous pathogens, cytokines, as well as many forms of tissue damage, can rapidly induce additional expression and translation of XOR [5]. Although XD is the predominant XOR form found in normal cells and tissues, XO appears to have an important role in cell and tissue injuries [5,6]. Various forms of stimuli induce the conversion of the XD to the XO form, presumably resulting in intensive synthesis of ROS and RNS [5,6].

XOR and uric acid are also secreted on the surface of (mucosal) epithelial tissues, thereby expanding their protective function extracellularly. Uric acid is found in nasal fluid, saliva and many other body fluids [14,30,31]. Uric acid is also released from the heart and the microvascular endothelium was identified as its site of formation [32]. XOR itself is constitutively released by

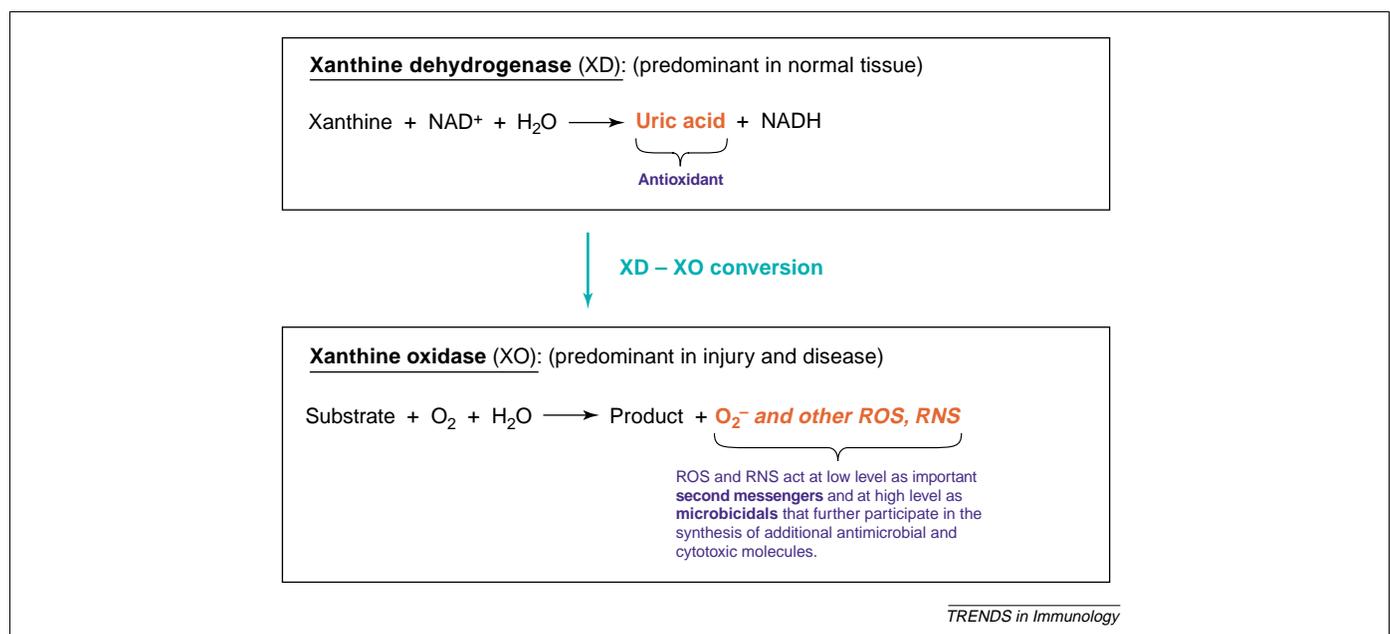


Fig. 1. XOR is a housekeeping enzyme with a role in purine catabolism, detoxification and the regulation of the cellular redox potential. XOR exists in two enzymatic forms, as a XD and as an XO. XD is the form that predominates in purine catabolism resulting in the synthesis of the antioxidant uric acid. The XO form is associated with the synthesis of large amount of ROS and RNS, which at low levels are important second messengers but at high levels have microbicidal action. The ability of XOR to rapidly convert from XD to XO under various forms of tissue injury and damage makes XOR an ideal component for a fast innate immune response. Abbreviations: NAD, nicotinamide adenine dinucleotide; ROS, reactive oxygen species; RNS reactive nitrogen species; XD, xanthine dehydrogenase; XO, xanthine oxidase; XOR, xanthine oxidoreductase.

pulmonary microvascular endothelial cells and found on the outside surface of endothelial plasma membranes of various organs [33–35]. More importantly, XOR is secreted by the lactating mammary gland and the small intestine for an antimicrobial purpose [19,36,37]. Although the mammary gland and milk provide immunological protection for the offspring, the intestine needs an intensive immune defense owing to its expansive epithelial surface, its multitude of villi and narrow invaginations, its exposure to dietary antigens and its proximity to the colon. XOR activity was detected in enterocytes and in the mucous of duodenum [38]. Within the small intestine, XOR was detected in Paneth cells, which are specialized epithelial cells that secrete various antimicrobial molecules in response to bacterial and other stimuli [39].

XOR is a mediator of infection and inflammation and interacts with NF- κ B and activator protein-1 (AP-1)

Inflammatory reactions are an important part of innate immunity, comprising a sequence of events induced by various forms of tissue damage and infection. The inflammatory reaction results in the expression of various cytokines and XOR is stimulated by interferon- γ (IFN- γ), IFN- α , tumor necrosis factor- α (TNF- α), interleukin-1 (IL-1) and IL-3; some of these factors also initiate the conversion from the XD to the XO form [5,6]. Endothelial XOR was suggested to trigger a microvascular inflammatory response, leading to recruitment and activation of circulating neutrophils and trapping of circulating pathogens [6]. The recruitment of neutrophils and the activation of macrophages are induced by ROS that upregulate cell adhesion molecules [40,41]. ROS from XOR and NADPH oxidase mediate the expression of endothelial-cell surface P-selectin, one of the first cell-surface molecules to be expressed on the endothelial cell in response to inflammation [42].

Throughout evolution, pathogen-induced stimulation of the innate immune system relies on numerous conserved Toll-like receptors (TLRs), which initiate intracellular signaling cascades that subsequently activate the transcription factors NF- κ B and AP-1 [43,44]. NF- κ B controls a wide variety of immune-inducible antimicrobial molecules, including ROS and RNS, as well as cytokines and co-stimulatory molecules [45]. Binding sites for NF- κ B and AP-1, as well as for response elements for cytokines, have been identified in the 5' UTR (untranslated region) of the human XOR gene and both NF- κ B and XOR are stimulated by bacterial lipopolysaccharids (LPS) *in vivo* [28,46,47]. This suggests that XOR is a downstream target of the TLR–NF- κ B pathway and is directly activated by NF- κ B and AP-1. Yet, both transcription factors are also stimulated by the redox potential of the cell and XOR might, therefore, also regulate the expression of NF- κ B and AP-1 [48,49]. The application of an inhibitor for XOR leads to a dose-dependent inhibition of NF- κ B during hepatic ischemia-reperfusion, and AP-1 is downregulated following the synthesis of NO [50,51]. Consequently, we suggest intensive cross talk between XOR, the TLR–NF- κ B pathway and the transcription factors NF- κ B and AP-1.

XOR participates in systemic protection and detoxification

XOR activity and uric acid are generally found in the blood plasma of many mammalian species and levels are particularly high during numerous disease states [5]. XOR might participate in the systemic antimicrobial response of the innate immune system by providing an oxidative defense mechanism in the blood. XOR is reported to have a defense role in meningitis, malaria and in trypanosomiasis [15,52,53]. Serum XOR is predominantly in the XO form, as a result of serum proteases [54]. XOR and the synthesis of ROS have also been implicated in the activation of the complement system, an important feature of the systemic innate immune system [55,56].

XOR is highly expressed in the liver for purine catabolism and nitrogen elimination as well as for systemic detoxification. XOR and AO are both important for the biotransformation of numerous toxic compounds [11]. Yet, the liver is also an organ for systemic bacterial scavenging, inactivation of bacterial products and inflammatory mediator clearance and production [57]. Liver Kupffer cells are a major source of ROS and proinflammatory cytokines that are produced in response to LPS, making the liver a key organ for the killing of circulating microorganisms [58]. ROS generated by both XOR and NADPH oxidase-dependent pathways are important for the systemic killing of *Candida parapsilosis* by Kupffer cells and the inhibition of XO activity leads to significant reduction of hepatic killing of various bacterial strains [59–62]. Furthermore, the liver produces various antimicrobial serum proteins that are released in response to infection or inflammation and generally are known as the acute phase proteins. The way cytokines stimulate XOR activity is similar to an acute phase response, and XOR might also be considered to participate in this feature of innate immunity [5,53]. Several other tissues and organs upregulate XOR in response to systemic infections, including the spleen, which is the major organ for pathogenic scavenging [28,63].

XOR has an important role in phagocytotic killing

Innate and adaptive immunity rely on the activity of phagocytotic cells that release numerous molecules, including ROS, for the intracellular destruction of phagocytosed microorganisms and for extracellular toxic effects on pathogens and tumor cells. Several enzymes are responsible for the synthesis of ROS in phagocytes and experiments suggest that XOR also participates in the innate immune response of phagocytotic cells [18,21]. Animals infected with *Staphylococcus aureus*, *Salmonella typhimurium*, *Plasmodium berghei*, *Schistosoma mansoni*, *Listeria*, various viruses and Ehrlich hyperdiploid carcinoma cells, all show a marked increase in XO activity in phagocytotic cells. Most of the XOR activity found in these infected animals is in the XO form. The administration of XOR inhibitors to infected animals leads to a decrease in the bactericidal activity of phagocytes and a linear dose-increase in mortality. Administration of purified XO or xanthine, a XOR substrate, to infected animals, results in significant protection against bacterial infections [61,64,65]. Furthermore, XOR contributes to

host defense against *Burkholderia cepacia* and phagocytic destruction of *Candida parapsilosis* by murine peritoneal macrophages [66,67].

XOR participates in the innate immune system throughout the animal kingdom

XOR has general cellular protective functions, presumably present in all eukaryotic and most prokaryotic cells. Yet, there is evidence that its role in systemic protection and innate immunity is also conserved. Because innate immunity is evolutionarily ancient and preceded adaptive immunity [68], the protective role of XOR in the innate immune system of lower organisms might be particularly crucial for their defense and survival. Invertebrates possess an efficient innate immune system composed of cellular and humoral elements. The humoral defense includes antimicrobial peptides, the cascades that regulate coagulation and melanization of hemolymph, and the production of ROS and RNS [69]. The involvement of cytotoxic ROS and RNS in innate immunity has been studied intensively in *Drosophila* and other insects, as well as in bivalve mollusks [21,24,70,71]. Uric acid is known to occur in large amounts in *Drosophila* and a decline of this antioxidant with age could be associated with the loss of antioxidant potential in aging [72]. The melanization reaction is performed by many arthropods as part of the innate immune system and triggered by injury and recognition of microbial cell-wall composition. Melanization leads to the synthesis and deposition of melanin in wounded areas, resulting in wound healing, synthesis of toxic products (including ROS and RNS), sequestration, encapsulation and killing of invading microorganisms [73,74]. XOR and ROS have been implicated in the synthesis of melanin and it has been postulated that the melanization of skin and other tissues is also an important component of the innate immune system of vertebrates [75].

In mammals, XOR, as well as AO, is involved in numerous hepatic and epithelial detoxification reactions. In insects, insecticide resistance and insecticide detoxification are important host defense mechanisms, resulting in the upregulation of various metabolic enzymes. These enzymes are also involved in anabolism and catabolism of a multitude of endogenous compounds and the protection against oxidative stress [76]. AO is coamplified with the *Culex* mosquito insecticide resistance-associated *esterases* and its 5' flanking region contains binding sites, known from other insecticide resistance genes [12,77]. As mentioned, the TLR–NF- κ B-signaling cascade has a central role in innate immunity throughout the animal kingdom and crosstalk between XOR, ROS and NF- κ B has been outlined.

The complex and versatile role of XOR, particularly in the mammalian innate immune system, could be linked to the unique rapid post-translational conversion from the XD to the XO form. Only mammalian XOR, but not XOR from chicken and *Drosophila*, can be converted from the XD to the XO form experimentally. XD is reversibly converted to XO by thiol oxidation or irreversibly by limited proteolysis [2,78]. The mechanism for this conversion might have evolved very recently. Phylogenetic

analysis of the presence of various oxidases demonstrate that although the XD form of XOR is present throughout all vertebrates except reptiles, the XO form is only found in mammals. XOR in its XD form is abundant in insects, although crustacea and gastropoda seem to have much higher levels of AO than XOR. Neither XOR nor AO activity was detectable in Porifera, Coelenterata, Platyhelminthes, Nematelminthes and Nemertea [79]. This suggests that the systemic protective role of XOR, and probably also of AO, is found predominantly in higher invertebrates and presumably, throughout all vertebrates, with a particular complex function in mammals. The possibility to rapidly, reversibly convert XOR from the XD to the XO form, might provide mammals with a powerful intrinsic switch, ideally suited as a component for a rapid, transient, innate immune response, independent of immunological memory. In general, most effector molecules of innate immunity need a fast conversion from a pre-existing, inactive to an active form, which is usually achieved by proteolytic processing of a precursor molecule [80]. Yet, other forms of post-translational activation of XOR could be of similar importance and the strong emphasis of the literature on the XD to XO conversion has been questioned [5,6]. XOR also exists in enzymatic inactive demolybdo and desulfo forms [5]. Experiments show that a desulpho–sulpho conversion of XOR is associated with an increase in XOR activity [5]. Moreover, activation of XOR by phosphorylation has been suggested [81]. It is possible that various forms of post-translational activation of XOR might act as switches for the complex role of XOR in numerous fast innate immune responses.

Conclusions

The ability of the multifunctional enzyme XOR to perform general detoxification reactions and additionally, to synthesize large amounts of the antioxidant uric acid, as well as ROS and RNS, makes it a versatile intra- and extra-cellular protective housekeeping enzyme and an important component of the innate immune system. XOR is involved in numerous features of mammalian innate immunity but, furthermore, its protective role appears to be evolutionarily conserved and downstream of the TLR–NF- κ B pathway. Because numerous features of the innate immune system include oxidative defense and XOR and elements of cellular oxidative protection predated the existence of the innate immune system, we suggest that XOR is a central molecule in the evolution and function of this ancient defense system (Fig. 2). Because the innate immune response generates many signals that co-stimulate the adaptive immune system, it is possible that XOR also has a role in the crosstalk between innate and adaptive immunity.

As mentioned, we recently discovered that XOR also has an essential role in milk fat droplet secretion from the lactating mammary gland. How XOR and the innate immune system contributed to the evolution of the mammary gland, milk and the entire class mammalia, will be discussed in another communication (C. Vorbach and M. Capecchi, unpublished).

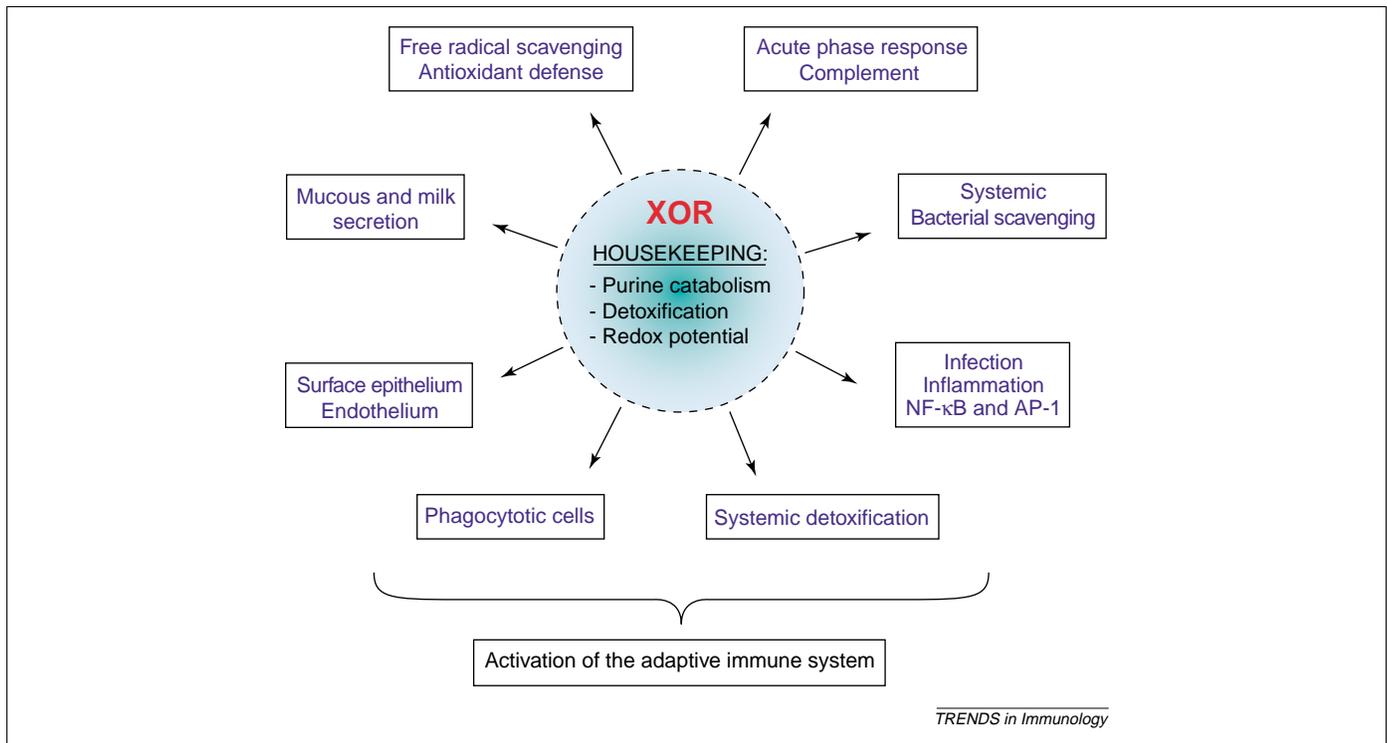


Fig. 2. XOR is an evolutionarily conserved housekeeping enzyme, important for purine catabolism, detoxification and the regulation of the cellular redox potential. Interestingly, the same protective functions are involved in multiple features of the innate immune system, suggesting that XOR is a central molecule in the evolution and function of this ancient defense system. Because innate immune responses generate many signals that co-stimulate the adaptive immune system it is possible that XOR also has a role in the crosstalk between innate and adaptive immunity. Abbreviations: AP-1, activator protein-1, XOR, xanthine oxidoreductase.

Acknowledgements

We would like to thank Lara Carroll for helping to prepare this manuscript.

References

- Bray, R.C. (1975) Molybdenum iron-sulfur flavin hydroxylases and related enzymes. In *The Enzymes* (Boyer, P.D., ed.), pp. 299–419, Academic Press
- Hille, R. and Nishino, T. (1995) Flavoprotein structure and mechanism. 4. Xanthine oxidase and xanthine dehydrogenase. *FASEB J.* 9, 995–1003
- Ichida, K. *et al.* (1997) Identification of two mutations in human xanthine dehydrogenase gene responsible for classical type I xanthinuria. *J. Clin. Invest.* 99, 2391–2397
- Vorbach, C. *et al.* (2002) The housekeeping gene xanthine oxidoreductase is necessary for milk fat droplet enveloping and secretion: gene sharing in the lactating mammary gland. *Genes Dev.* 16, 3223–3235
- Harrison, R. (2002) Structure and function of xanthine oxidoreductase: where are we now? *Free Radic. Biol. Med.* 33, 774–797
- Meneshian, A. and Bulkley, G.B. (2002) The physiology of endothelial xanthine oxidase: from urate catabolism to reperfusion injury to inflammatory signal transduction. *Microcirculation* 9, 161–175
- Squadrito, G.L. *et al.* (2000) Reaction of uric acid with peroxynitrite and implications for the mechanism of neuroprotection by uric acid. *Arch. Biochem. Biophys.* 376, 333–337
- Sheehan, D. *et al.* (2001) Structure, function and evolution of glutathione transferases: implications for classification of non-mammalian members of an ancient enzyme superfamily. *Biochem. J.* 360, 1–16
- Gus'kov, E.P. *et al.* (2002) Allantoin as a free-radical scavenger. *Dokl. Biochem. Biophys.* 383, 105–107
- Droge, W. (2002) Free radicals in the physiological control of cell function. *Physiol. Rev.* 82, 47–95
- Beedham, C. (2002) Molybdenum hydroxylases. In *Enzyme Systems that Metabolise Drugs and Other Xenobiotics* (Ionnides, C., ed.), pp. 147–187, John Wiley
- Garattini, E. *et al.* (2003) Mammalian molybdo-flavoenzymes, an expanding family of proteins: structure, genetics, regulation, function and pathophysiology. *Biochem. J.* 372, 15–32
- Baeuerle, P.A. and Henkel, T. (1994) Function and activation of NF-κB in the immune system. *Annu. Rev. Immunol.* 12, 141–179
- Becker, B.F. (1993) Towards the physiological function of uric acid. *Free Radic. Biol. Med.* 14, 615–631
- Christen, S. *et al.* (2001) Marked elevation in cortical urate and xanthine oxidoreductase activity in experimental bacterial meningitis. *Brain Res.* 900, 244–251
- Nagler, R.M. *et al.* (2002) Characterization of the differentiated antioxidant profile of human saliva. *Free Radic. Biol. Med.* 32, 268–277
- Babior, B.M. *et al.* (1975) Biological defense mechanisms. Evidence for the participation of superoxide in bacterial killing by xanthine oxidase. *J. Lab. Clin. Med.* 85, 235–244
- Rosen, G.M. *et al.* (1995) Free radicals and phagocytic cells. *FASEB J.* 9, 200–209
- Hancock, J.T. *et al.* (2002) Antimicrobial properties of milk: dependence on presence of xanthine oxidase and nitrite. *Antimicrob. Agents Chemother.* 46, 3308–3310
- Granger, D.N. *et al.* (1981) Superoxide radicals in feline intestinal ischemia. *Gastroenterology* 81, 22–29
- Anderson, R.S. (2001) Reactive oxygen species and antimicrobial defenses of invertebrates: a bivalve model. *Adv. Exp. Med. Biol.* 484, 131–139
- Godber, B.L. *et al.* (2000) Reduction of nitrite to nitric oxide catalyzed by xanthine oxidoreductase. *J. Biol. Chem.* 275, 7757–7763
- Brunelli, L. *et al.* (1995) The comparative toxicity of nitric oxide and peroxynitrite to *Escherichia coli*. *Arch. Biochem. Biophys.* 316, 327–334
- Nappi, A.J. and Vass, E. (2001) Cytotoxic reactions associated with insect immunity. *Adv. Exp. Med. Biol.* 484, 329–348
- Taibi, G. *et al.* (2001) Xanthine oxidase catalyzes the synthesis of retinoic acid. *J. Enzyme Inhib.* 16, 275–285
- Jarasch, E.D. *et al.* (1981) Localization of xanthine oxidase in mammary-gland epithelium and capillary endothelium. *Cell* 25, 67–82
- Kooij, A. *et al.* (1992) High levels of xanthine oxidoreductase in rat endothelial, epithelial and connective tissue cells. A relation between

- localization and function? *Virchows Arch. B Cell Pathol. Incl. Mol. Pathol.* 62, 143–150
- 28 Kurosaki, M. *et al.* (1995) Tissue- and cell-specific expression of mouse xanthine oxidoreductase gene *in vivo*: regulation by bacterial lipopolysaccharide. *Biochem. J.* 306, 225–234
- 29 Linder, N. *et al.* (1999) Cellular expression of xanthine oxidoreductase protein in normal human tissues. *Lab. Invest.* 79, 967–974
- 30 Peden, D.B. *et al.* (1990) Uric acid is a major antioxidant in human nasal airway secretions. *Proc. Natl. Acad. Sci. U. S. A.* 87, 7638–7642
- 31 Kastenbauer, S. *et al.* (2001) Experimental meningitis in the rat: protection by uric acid at human physiological blood concentrations. *Eur. J. Pharmacol.* 425, 149–152
- 32 Becker, B.F. *et al.* (1989) Uric acid as radical scavenger and antioxidant in the heart. *Pflugers Arch.* 415, 127–135
- 33 Partridge, C.A. *et al.* (1992) Pulmonary microvascular endothelial cells constitutively release xanthine oxidase. *Arch. Biochem. Biophys.* 294, 184–187
- 34 Vickers, S. *et al.* (1998) Immunoaffinity localization of the enzyme xanthine oxidase on the outside surface of the endothelial cell plasma membrane. *Surgery* 124, 551–560
- 35 Rouquette, M. *et al.* (1998) Xanthine oxidoreductase is asymmetrically localised on the outer surface of human endothelial and epithelial cells in culture. *FEBS Lett.* 426, 397–401
- 36 Collins, R.A. *et al.* (1988) Histochemical localization and possible antibacterial role of xanthine oxidase in the bovine mammary gland. *J. Dairy Res.* 55, 25–32
- 37 Stevens, C.R. *et al.* (2000) Antibacterial properties of xanthine oxidase in human milk. *Lancet* 356, 829–830
- 38 Van Den Munckhof, R.J. *et al.* (1995) Ultrastructural localization of xanthine oxidase activity in the digestive tract of the rat. *Histochem. J.* 27, 897–905
- 39 Morita, Y. *et al.* (2001) Identification of xanthine dehydrogenase/xanthine oxidase as a rat paneth cell zinc-binding protein. *Biochim. Biophys. Acta* 1540, 43–49
- 40 Kurose, I. and Granger, D.N. (1994) Evidence implicating xanthine oxidase and neutrophils in reperfusion-induced microvascular dysfunction. *Ann. N. Y. Acad. Sci.* 723, 158–179
- 41 Granger, D.N. and Kubes, P. (1994) The microcirculation and inflammation: modulation of leukocyte-endothelial cell adhesion. *J. Leukoc. Biol.* 55, 662–675
- 42 Takano, M. *et al.* (2002) Rapid upregulation of endothelial P-selectin expression via reactive oxygen species generation. *Am. J. Physiol. Heart Circ. Physiol.* 283, H2054–H2061
- 43 Kopp, E.B. and Medzhitov, R. (1999) The Toll-receptor family and control of innate immunity. *Curr. Opin. Immunol.* 11, 13–18
- 44 Silverman, N. and Maniatis, T. (2001) NF- κ B signaling pathways in mammalian and insect innate immunity. *Genes Dev.* 15, 2321–2342
- 45 Zhang, G. and Ghosh, S. (2001) Toll-like receptor-mediated NF- κ B activation: a phylogenetically conserved paradigm in innate immunity. *J. Clin. Invest.* 107, 13–19
- 46 Xu, P. *et al.* (1996) Molecular cloning and characterization of the human xanthine dehydrogenase gene (XDH). *Genomics* 34, 173–180
- 47 Hassoun, P.M. *et al.* (1998) Upregulation of xanthine oxidase by lipopolysaccharide, interleukin-1, and hypoxia. Role in acute lung injury. *Am. J. Respir. Crit. Care Med.* 158, 299–305
- 48 Meyer, M. *et al.* (1994) Regulation of the transcription factors NF- κ B and AP-1 by redox changes. *Chem. Biol. Interact.* 91, 91–100
- 49 Forman, H.J. *et al.* (2002) Redox signaling. *Mol. Cell. Biochem.* 234, 49–62
- 50 Matsui, N. *et al.* (2000) Xanthine oxidase-derived reactive oxygen species activate nuclear factor- κ B during hepatic ischemia in rats. *Jpn. J. Pharmacol.* 84, 363–366
- 51 Natarajan, R. *et al.* (2001) Nitric oxide suppresses IL-8 transcription by inhibiting c-Jun N-terminal kinase-induced AP-1 activation. *Exp. Cell Res.* 266, 203–212
- 52 Bungener, W. (1974) Influence of allopurinol on the multiplication of rodent malaria parasites. *Tropenmed. Parasitol.* 25, 309–312
- 53 Wang, J. *et al.* (2002) Serum xanthine oxidase: origin, regulation, and contribution to control of trypanosome parasitemia. *Antioxid. Redox Signal.* 4, 161–178
- 54 Kooij, A. *et al.* (1994) Conversion of xanthine dehydrogenase into xanthine oxidase in rat liver and plasma at the onset of reperfusion after ischemia. *Hepatology* 19, 1488–1495
- 55 Turnage, R.H. *et al.* (1994) Complement activation by the hydroxyl radical during intestinal reperfusion. *Shock* 2, 445–450
- 56 Tanhehco, E.J. *et al.* (2000) Free radicals upregulate complement expression in rabbit isolated heart. *Am. J. Physiol. Heart Circ. Physiol.* 279, H195–H201
- 57 Cerra, F.B. (1987) Hypermetabolism, organ failure, and metabolic support. *Surgery* 101, 1–14
- 58 Klein, A. *et al.* (1994) Quantitative discrimination of hepatic reticuloendothelial clearance and phagocytic killing. *J. Leukoc. Biol.* 55, 248–252
- 59 Tubaro, E. *et al.* (1980) Liver xanthine oxidase increase in mice in three pathological models. A possible defence mechanism. *Biochem. Pharmacol.* 29, 1939–1943
- 60 Klein, A.S. *et al.* (1996) Allopurinol: discrimination of antioxidant from enzyme inhibitory activities. *Free Radic. Biol. Med.* 21, 713–717
- 61 Umezawa, K. *et al.* (1997) Induction of nitric oxide synthesis and xanthine oxidase and their roles in the antimicrobial mechanism against *Salmonella typhimurium* infection in mice. *Infect. Immun.* 65, 2932–2940
- 62 Potoka, D.A. *et al.* (1998) Endothelial cells potentiate oxidant-mediated Kupffer cell phagocytic killing. *Free Radic. Biol. Med.* 24, 1217–1227
- 63 Chinnaiyan, A.M. *et al.* (2001) Molecular signatures of sepsis: multiorgan gene expression profiles of systemic inflammation. *Am. J. Pathol.* 159, 1199–1209
- 64 Crosby, P.F. *et al.* (1969) Liver xanthine oxidase activity of mice infected with *Schistosoma mansoni*. *J. Parasitol.* 55, 673
- 65 Tubaro, E. *et al.* (1980) Xanthine oxidase increase in polymorphonuclear leucocytes and macrophages in mice in three pathological situations. *Biochem. Pharmacol.* 29, 1945–1948
- 66 Takao, S. *et al.* (1996) Role of reactive oxygen metabolites in murine peritoneal macrophage phagocytosis and phagocytic killing. *Am. J. Physiol.* 271, C1278–C1284
- 67 Segal, B.H. *et al.* (2000) Xanthine oxidase contributes to host defense against *Burkholderia cepacia* in the p47(phox^{-/-}) mouse model of chronic granulomatous disease. *Infect. Immun.* 68, 2374–2378
- 68 Fearon, D.T. and Locksley, R.M. (1996) The instructive role of innate immunity in the acquired immune response. *Science* 272, 50–53
- 69 Lavine, M.D. and Strand, M.R. (2002) Insect hemocytes and their role in immunity. *Insect Biochem. Mol. Biol.* 32, 1295–1309
- 70 Nappi, A.J. *et al.* (1995) Superoxide anion generation in *Drosophila* during melanotic encapsulation of parasites. *Eur. J. Cell Biol.* 68, 450–456
- 71 Nappi, A.J. *et al.* (2000) Nitric oxide involvement in *Drosophila* immunity. *Nitric Oxide* 4, 423–430
- 72 Massie, H.R. *et al.* (1991) Uric acid content of *Drosophila* decreases with aging. *Exp. Gerontol.* 26, 609–614
- 73 Lee, W.J. *et al.* (1998) Molecular cloning and chromosomal localization of a prophenoxyoxidase cDNA from the malaria vector *Anopheles gambiae*. *Insect Mol. Biol.* 7, 41–50
- 74 Soderhall, K. and Cerenius, L. (1998) Role of the prophenoxyoxidase-activating system in invertebrate immunity. *Curr. Opin. Immunol.* 10, 23–28
- 75 Mackintosh, J.A. (2001) The antimicrobial properties of melanocytes, melanosomes and melanin and the evolution of black skin. *J. Theor. Biol.* 211, 101–113
- 76 Ranson, H. *et al.* (2002) Evolution of supergene families associated with insecticide resistance. *Science* 298, 179–181
- 77 Coleman, M. *et al.* (2002) Molecular characterization of the amplified aldehyde oxidase from insecticide resistant *Culex quinquefasciatus*. *Eur. J. Biochem.* 269, 768–779
- 78 Amaya, Y. *et al.* (1990) Proteolytic conversion of xanthine dehydrogenase from the NAD-dependent type to the O₂-dependent type. Amino acid sequence of rat liver xanthine dehydrogenase and identification of the cleavage sites of the enzyme protein during irreversible conversion by trypsin. *J. Biol. Chem.* 265, 14170–14175
- 79 Wurzinger, K.H. and Hartenstein, R. (1974) Phylogeny and correlations of aldehyde oxidase, xanthine oxidase, xanthine dehydrogenase and peroxidase in animal tissues. *Comp. Biochem. Physiol. B* 49, 171–185
- 80 Boman, H.G. (2000) Innate immunity and the normal microflora. *Immunol. Rev.* 173, 5–16
- 81 Kayyali, U.S. *et al.* (2001) Phosphorylation of xanthine dehydrogenase/oxidase in hypoxia. *J. Biol. Chem.* 276, 14359–14365